Transcription, Chromatin, and Epigenetics:

Ligand-dependent Corepressor LCoR Is an Attenuator of Progesterone-regulated Gene Expression

Ana Palijan, Isabelle Fernandes, Mark Verway, Maria Kourelis, Yolande Bastien, Luz E. Tavera-Mendoza, Aaron Sacheli, Veronique Bourdeau, Sylvie Mader and John H. White

doi: 10.1074/jbc.M109.051201 originally published online September 10, 2009

Access the most updated version of this article at doi: 10.1074/jbc.M109.051201

Find articles, minireviews, Reflections and Classics on similar topics on the JBC Affinity Sites.

Alerts:
- When this article is cited
- When a correction for this article is posted

Click here to choose from all of JBC's e-mail alerts

Supplemental material:
http://www.jbc.org/content/suppl/2009/09/10/M109.051201.DC1.html

This article cites 54 references, 29 of which can be accessed free at
http://www.jbc.org/content/284/44/30275.full.html#ref-list-1
Ligand-dependent Corepressor LCoR Is an Attenuator of Progesterone-regulated Gene Expression

Ana Palijan1, Isabelle Fernandes1,2, Mark Verway1, Maria Koulalis1, Yolande Bastien3, Luz E. Tavares-Mendoza6,3, Aaron Sacheli1, Veronique Bourdeau4, Sylvie Mader1, and John H. White1,3,5

From the Departments of 1Physiology and 2Medicine, McGill University, Montreal, Quebec H3G 1Y6, Canada, the 3Department of Adult Oncology, Dana-Farber Cancer Institute, Harvard Medical School, Boston, Massachusetts 02115, and the 4Institute for Research in Immunology and Cancer and Biochemistry Department, University of Montreal, Montreal, Quebec H3C 3J7, Canada

Ligand-dependent corepressor LCoR interacts with the progesterone receptor (PR) and estrogen receptor ERα in the presence of hormone. LCoR contains tandem N-terminal PXDLS motifs that recruit C-terminal-binding protein (CtBP) corepressors as well as a C-terminal helix-turn-helix (HTH) domain. Here, we analyzed the function of these domains in coregulation of PR- and ERα-regulated gene expression. LCoR and CtBP1 colocalize in nuclear bodies that also contain CtBP-interacting protein CtIP and polycomb group repressor complex marker BMI1. Coexpression of CtBP1 in MCF7 or T47D breast cancer cells augmented corepression by LCoR, whereas coexpression of CtIP did not, consistent with direct interaction of LCoR with CtBP1, but not CtIP. The N-terminal region containing the PXDLS motifs is necessary and sufficient for CtBP1 recruitment and essential for full corepression. However, LCoR function was also strongly dependent on the helix-turn-helix domain, as its deletion completely abolished corepression. LCoR, CtBP, and CtIP were recruited to endogenous PR- and ERα-stimulated genes in a hormone-dependent manner. Similarly, LCoR was recruited to estrogen-repressed genes, whereas hormone treatment reduced CtBP1 binding. Small interfering RNA-mediated knockdown of LCoR or CtBP1 augmented expression of progesterone- and estrogen-stimulated reporter genes as well as endogenous progesterone-stimulated target genes. In contrast, their ablation had gene-specific effects on ERα-regulated transcription that generally led to reduced gene expression. Taken together, these results show that multiple domains contribute to LCoR function. They also reveal a role for LCoR and CtBP1 as attenuators of progesterone-regulated transcription but suggest that LCoR and CtBP1 can act to enhance transcription of some genes.

The progesterone receptor (PR)6 and estrogen receptor α (ERα) are members of the nuclear receptor superfamily of hormone-regulated transcription factors whose functions have been implicated in a broad range of physiological responses (1–7). During activation of gene transcription, agonist-bound nuclear receptors recruit so-called coactivators such as p160 proteins (2, 8–10). Coactivators act in part by functioning as histone acetyltransferases (HATs) or by recruiting HAT activity to target promoters. Notably, screens designed to identify coactivators revealed that agonist-bound nuclear receptors also recruit proteins with corepressor properties, such as nuclear receptor-interacting protein 1 (RIP140/ NRP1) (11–14) or LCoR (11, 15). We identified LCoR as a protein that interacted with the estrogen-bound ligand binding domain of ERα. Transiently expressed LCoR repressed hormone-dependent transactivation by a range of nuclear receptors. This included particularly efficacious repression of progesterone-stimulated transcription. LCoR transcripts are detectable as early as the two-cell stage of embryonic development, and the protein is robustly expressed in numerous fetal and adult tissues, including progesterone and estrogen target tissues such as breast, endometrium, and placenta (15).

LCoR acts as a scaffold for both histone deacetylases (HDACs) and C-terminal binding protein (CtBP) corepressors and functions by both HDAC-dependent and -independent mechanisms in a receptor-specific manner. For example, whereas the HDAC inhibitor trichostatin A abolished LCoR-dependent corepression of estrogen-stimulated reporter gene expression, PR-stimulated expression was largely resistant to trichostatin A (15). LCoR contains two adjacent sequences, (P/V)XLDLX(K/R) or (P/V)XLDLXXK, near its N terminus that correspond to extended PXDLS consensus motifs recognized by CtBPs (13). CtBP1 was identified as a protein that bound the C-terminal region of human adenosviral E1A (16) and is the

6 The abbreviations used are: PR, progesterone receptor; ChIP, chromatin immunoprecipitation; CtBP, C-terminal binding protein; CtIP, CtBP-interacting protein; CyPB, cyclophilin B; E2, estradiol; ERα, estrogen receptor α; ERE, estrogen response element; HDAC, histone deacetylase; HTH, helix-turn-helix; LCoR, ligand-dependent corepressor; MBLK-1, honeybee MBlk-1-related factor; P4, progesterone; PRC1, polycomb group repressor complexes; pS2, trefoil factor 1; PSQ, pipsqueak motif; siRNA, small interfering RNA; qRT, quantitative real-time; SGK3, serum/glucocorticoids-regulated kinase 3; BMP7, bone morphogenetic protein 7; DEMM, Dulbecco’s modified Eagle’s medium; BSA, bovine serum albumin; GST, glutathione S-transferase; IGFBP1, insulin-like growth factor binding protein 1; CYP26B1, cytochrome P450, family 26, subfamily b, polypeptide 1.

Received for publication, July 31, 2009, and in revised form, September 8, 2009. Published, JBC Papers in Press, September 10, 2009, DOI 10.1074/jbc.M109.051201
LCoR Is a Corepressor of Progesterone-regulated Gene Expression

founder of a well conserved family of proteins (17–19). Highly homologous CtBP2 was identified from EST databases (20). Remarkably, CtBP1 bind NAD(H) and are related to 2-hydroxy acid dehydrogenases, and CtBP1 has weak dehydrogenase activity. In addition, dinucleotide binding stimulates protein oligomerization activity and corepression.

CtBPs interact directly with several transcriptional coregulatory proteins, many of which share the PXDLS motifs described above (13, 17, 21). For example, a screen for CtBP cofactors identified CtBP interacting protein (CtIP) (22), which also binds BRCA1 and retinoblastoma gene product (Rb) tumor suppressor proteins (23, 24). Like LCoR, CtBPs can function by HDAC-dependent or -independent mechanisms depending on the promoter tested (17). They are components of several multisubunit assemblies, including polycomb repressor PRC1 complexes (25–27). Targeted ablation of CtBP1 or -2 expression in mice revealed that the two proteins play important and overlapping roles in mouse development (19, 28).

In addition to its N-terminal CtBP interaction motifs and a central HDAC binding domain (29), LCoR contains a C-terminal helix-turn-helix (HTH) domain that is homologous to motifs encoded by the Eip93F (CG18389 gene product from Drosophila melanogaster) and MBLK-1 genes of Drosophila and Honeybee (Apis mellifera) (30), respectively. The LCoR HTH domain also bears 35% homology to pipsqueak motifs (PSQ). PSQ motifs are repeated four times in the DNA binding region of the Drosophila pipsqueak transcription factor, which plays a role in gene silencing (31). Multiple repeats of the domain are required for PSQ DNA binding (31), and mutation of one of the two HTH motifs in the MBLK-1 gene strongly reduced site-specific DNA binding (30). The PSQ domain is homologous to unique motifs found in a number of prokaryotic and eukaryotic proteins that interact with DNA, such as recombinases (31, 32), raising the possibility that LCoR itself may interact with DNA. Other studies have shown that HTH domains can function in protein-protein interactions, where the HTH motif, combined with other domains, can induce formation of multisubunit complexes (33). Such proteins with multidomains can act as scaffolds between the basal transcription machinery and transcription factors (33).

The accompanying paper showed that LCoR recruits HDAC6 through a central domain (29). In this study we have analyzed the roles of domains controlling recruitment of CtBPs and the C-terminal HTH motif in corepression by LCoR. We were primarily interested in determining the roles of these domains in corepression of the PR, as our previous study showed that the efficacious corepression of PR-driven gene reporter gene expression by LCoR appeared to be largely insensitive to HDAC inhibition (15). We find that both the PXDLS motifs and the HTH domain are required for corepression of both the PR and ERα, as disruption of either region markedly attenuated LCoR function. LCoR colocalizes with CtBP1 and CtIP in nuclear foci, and CtBP1 is corecruited with LCoR to PR and ERα target genes in a hormone-dependent manner. Ablation of LCoR and/or CtBP1 had gene-specific effects on ERα-regulated genes and generally led to reduced target gene expression.

**EXPERIMENTAL PROCEDURES**

Antibodies—A rabbit polyclonal antipeptide antibody was raised against LCoR amino acids 20–36 (QDPSPQPNSTKNSEQ-LPKA) fused to keyhole limpet hemocyanin and purified over a peptide affinity column (Bethyl Laboratories, Montgomery, TX). Rabbit polyclonal α-CtBP (sc-11390), goat polyclonal α-CtBP1 (sc-5963), goat polyclonal α-CtIP (sc-5970), goat polyclonal α-Bmi1 (sc-8906), rabbit polyclonal α-Bmi1 (sc-10745), mouse monoclonal glyceraldehyde-3-phosphate dehydrogenase (sc-69778), rabbit polyclonal ERα (sc-543), rabbit polyclonal OCTA-Probe (sc-807), goat anti-mouse IgG (sc-2005), goat anti-rabbit IgG (sc-2004), normal mouse IgG (sc-2025), normal rabbit IgG (sc-2207), and protein G Plus-agarose (sc-2001) and protein G Plus-agarose (sc-2002) were from Santa Cruz Biotechnology (Santa Cruz, CA). Cy3-donkey polyclonal α-goat (705-165-147) and Cy2-goat polyclonal α-rabbit (711-225-152), Cy3-donkey polyclonal α-rabbit (711-165-152), and Cy2-donkey polyclonal α-mouse (715-225-150) were purchased from Jackson ImmunoResearch (West Grove, PA). Mouse monoclonal α-FLAG M2 (F3165) and α-FLAG M2 horseradish peroxidase (HRP)-conjugate (A-8592), monoclonal α-rabbit HRP conjugate (A2074), and rabbit polyclonal α-goat HRP conjugate (A5420) were from Sigma. Rabbit polyclonal CtBP1 (07-306) was purchased from Millipore (Temecula, CA). Rabbit polyclonal LCoR (sc-2005-44018) was purchased from GenWay Biotech (San Diego, CA). Rabbit polyclonal PR (ab68195) and rabbit polyclonal CtIP (ab70163) were purchased from Abcam (Cambridge, MA).

Recombinant Plasmids—PSG5/LCoR, FLAG-LCoR/pcDNA3.1 and LCoR derivatives mutagenized in the CtBP binding motifs, PLDLTVR (LCoR amino acids 64–70; m1) and VLDLSTK (LCoR amino acids 82–88; m2), and the double mutant (m1m2) have been described (3). cDNAs mutated in the CtBP binding motifs were subcloned downstream of FLAG in pCDNA3.1. FLAG-LCoRΔHTH/pcDNA3.1 was made using the QuikChange mutagenesis kit (Stratagene 200518, La Jolla, CA) as per the manufacturer’s instructions. Primers were designed to delete amino acids 350–395 from LCoR. The new construct was sequenced to confirm proper deletion LCoR.

Cell Culture and Transfections—All cells were cultured under the recommended conditions. For immunocytochemistry, MCF7 cells grown on collagen IV-treated microscope slides in 6-well plates in DMEM supplemented with 10% FBS. Cells were prepared for immunocytochemistry as described below. For immunoprecipitation of tagged proteins, MCF7 cells in 100-mm dishes were transfected with 10 μl of Lipofectamine containing 10 μg of pSG5 vectors containing FLAG-LCoR, FLAG-m1, FLAG-m2, or FLAG-m1 + m2. For analysis of the effects of CtBP1 and CtIP on LCoR corepression, MCF7 cells (60–70% confluent) were grown in DMEM without phenol red supplemented with 10% FBS on 6-well plates. Cells were transfected in medium without serum (Opti-MEM (Invitrogen)). The amounts of expression vectors were as follows: 100 ng of ERα or human PR.
expression vectors (as indicated), 100 ng of LCoR/pcDNA3.1, 250 ng of ERE3-TATA/pXP2 or pGRE5/pXP2 reporter plasmid, 250 ng of internal control vector pCMV-H9252gal. Quantities of expression vectors used are indicated in the legends to Figs. 3 and 5. The medium was replaced 18 h after transfection by a medium containing charcoal-stripped serum and ligand (10 nM) for 30 h as indicated. For the Luciferase reporter assay, cells were harvested in 250 l of reporter lysis buffer (Promega).

Immunocytochemistry and Immunoprecipitations—Cells were cultivated on collagen IV-treated microscope slides in 6-well plates, fixed with 2% paraformaldehyde for 15 min at room temperature, washed (3×) with phosphate-buffered saline and permeabilized with 0.2% Triton X-100, 5% BSA, 10% horse serum in phosphate-buffered saline. MCF7 cells were then incubated with α-LCoR (1:500) and goat polyclonal antibodies against CtBP1, CtIP, or Bmi1 (1:50) in buffer B (0.2% Triton X-100, 5% BSA 1 ml of LB, 48 h after transfection. Supernatants were cleared and incubated overnight with 4 μg of αCtBP or αCtIP or polyclonal rabbit αBMI1 or control rabbit or goat IgG at 4 °C overnight followed by 2 h of incubation at 4 °C with protein A-agarose (for αCtBP, αBmi1, control rabbit IgG) or protein A+G-agarose (for αCtIP or control goat IgG). Beads were washed (3×) with LB. Bound immunocomplexes were boiled in Laemmli buffer, separated by 10% SDS/PAGE, blotted on polyvinylidene difluoride membranes with α-LCoR (1/1000), α-CtBP1, α-CtBP2, α-CtIP, or α-BMI1 (1:100), and detected by enhanced chemiluminescence (PerkinElmer Life Sciences). For immunoprecipitation of tagged proteins, transfected MCF7 cells were lysed for 30 min at 4 °C in phosphate-buffered saline) for 1 h at room temperature. Cells were washed (3×) with phosphate-buffered saline and incubated with goat anti-rabbit-Cy2 and donkey anti-goat Cy3 (1:300) in buffer B for 1 h at room temperature. Slides were mounted with Immuno-Fluoro Mounting Medium (ICN, Aurora, Ohio) and visualized using a Zeiss LSM 510 confocal microscope.

For immunoprecipitation of endogenous CtBP, CtIP, or Bmi1, MCF7 cells in 150-mm dishes were lysed for 3 min at 4 °C in 1 ml of LB (150 mM NaCl, 10 mM Tris-HCl, pH 7.4, 0.2 mM sodium orthovanadate, 1 mM EDTA, 1 mM EGTA, 1% Triton X-100, 0.5% IGEPAL CA-630, protease inhibitor mixture). Cell debris was pelleted by centrifugation (14,000 rpm, 5 min), and proteins were immunoprecipitated with 4 μg of αCtBP or αCtIP or polyclonal rabbit αBMI1 or control rabbit or goat IgG at 4 °C overnight followed by 2 h of incubation at 4 °C with protein A-agarose (for αCtBP, αBmi1, control rabbit IgG) or protein A+G-agarose (for αCtIP or control goat IgG). Beads were washed (3×) with LB. Bound immunocomplexes were boiled in Laemmli buffer, separated by 10% SDS/PAGE, blotted on polyvinylidene difluoride membranes with α-LCoR (1/1000), α-CtBP1, α-CtBP2, α-CtIP, or α-BMI1 (1:100), and detected by enhanced chemiluminescence (PerkinElmer Life Sciences). For immunoprecipitation of tagged proteins, transfected MCF7 cells were lysed for 30 min at 4 °C in phosphate-buffered saline.)
siRNA Knockdowns—siRNAs were purchased from Thermo Scientific Dharmacon (Lafayette, CO). The following ON-TARGETplus SMART pool siRNAs were used: LCoR (L-026303-00), CtBP1 (L-008609-00), CyPB (D-001820-10), and non-targeting (D-001818-10). siRNAs were resuspended per the manufacturer's instructions. Transfections were done in 6-well plates as described previously. Lipofectamine 2000 (10 µl) was used as the transfection reagent. DMEM phenol-free with 10% stripped FBS was added 12 h after transfection. For Western blot analysis, cells were collected 48 h after transfection. Luciferase reporter assays after siRNA knockdowns were performed as follows. 100 ng of ERα expression vector and 250 ng of ERE3-TATA-pXP2 vector were transfected with the corresponding siRNA. DMEM phenol-free with 10% stripped FBS was added 12 h after transfection. Ligand was added 36 h after transfection, and cells were collected 24 h later. Luciferase activity was measured as previously described.

RNA Isolation, cDNA Synthesis, and Quantitative Real-time (qRT)-PCR—Cells were grown in 100-mm dishes. Media were replaced with charcoal-stripped medium containing ligand. Total RNA was extracted with TRIzol reagent. cDNA synthesis was performed with iScript cDNA synthesis kit (Bio-Rad) according to the instructions of the manufacturer. The MiniOpticon real-time PCR system with the iQ SYBR Green Supermix (Bio-Rad) was used for qRT-PCR expression analysis of target genes. The program used was as follows: 1) incubation at 94 °C for 60 s, 2) incubation at 95 °C for 20 s, 3) incubation at 60 °C for 30 s (decreasing temperature by 1° per cycle), 4) incubation at 72 °C for 30 s, 5) plate reading, 6) repetition from step 2 five more times, 7) incubation at 95 °C for 20 s, 8) incubation at 57.5 °C for 30 s, 9) incubation at 72 °C for 30 s, 10) plate reading, 11) repetition from step 7 thirty-five more times, 12) performance of melting curve and end. Results were normalized to β-actin mRNA expression. For qRT-PCR primers sequences, please refer to supplemental Table 2.
RESULTS

Association of LCoR with CtBP1 and CtIP—Colocalization of LCoR with CtBP1 in MCF7 cells was confirmed by immunocytochemical analyses (Fig. 1A). Both proteins were broadly distributed in the nucleus and were also concentrated in discrete nuclear bodies. A similar colocalization of CtBP2 and LCoR was also observed (data not shown). Given the extensive overlap of CtBPs and LCoR, we determined whether LCoR colocalized with CtIP, which was identified as a CtBP-interacting protein containing an extended PXDLS motif (22). Similar to results obtained with CtBP, CtIP and LCoR showed strongly overlapping patterns of expression concentrated in discrete nuclear bodies (Fig. 1B). In addition, endogenous LCoR coimmunoprecipitates with antibodies directed against either CtBPs or CtIP (Fig. 1C).

Other studies have shown that CtIP interacts directly with the retinoblastoma gene product (22) and link CtBP1 and Rb to polycomb group repressor complexes (26, 36). PRC1 complexes form large foci containing numerous factors, including BMI1 polycomb ring finger oncogene (BMI1), visible as discrete nuclear structures (37). Indeed, we found that BMI1 and LCoR coimmunoprecipitated and colocalized in nuclear bodies (supplemental Fig. 1). Taken together, these studies show that LCoR extensively colocalizes with CtBP1 and CtIP in the nucleus, including in PRC1 complexes.

The specificity of the interactions of LCoR with CtBP1 were further analyzed in vitro using glutathione S-transferase (GST) fused to a series of C-terminal deletion mutants of LCoR or LCoR mutant m1m2, which lacks the tandem PXDLS motifs (Fig. 2A). CtBP1 bound to full-length LCoR (Fig. 2B, top) and all C-terminal deletion mutants (Fig. 2B, bottom) but not to LCoR mutant m1m2 (Fig. 2A). Moreover, tagged LCoR mutated in either one of the two PXDLS motifs coimmunoprecipitated with endogenous CtBPs from extracts of MCF7 cells (m1, amino acids 64–70 deleted; m2, amino acids 82–88 deleted; Fig. 2C, bottom panel). In contrast, no coimmunoprecipitation was observed in cells express-
LCoR Is a Corepressor of Progesterone-regulated Gene Expression

A  IGFBP1 promoter
    -910          -50
    HRE   +1
Target region Non-Target region

B  0'  45'  0'  45'
P4
Input
IgG
PR
Flag-LCoR
CtBP1
CtIP

MCF7  T47D

Input
IgG
PR
Flag-LCoR
Non-Target control

E2
0'  30'
Input
IgG
PR
Flag-LCoR

MCF7

C  Input
IgG
Flag reChIP PR
Flag reChIP CtBP1

D  0'  30'
Input
IgG
ERα
Flag-LCoR
CtBP1
CtIP

Non-Target control

E  Input
IgG
Flag reChIP ERα
Flag reChIP CtBP1

pS2
LCoR Is a Corepressor of Progesterone-regulated Gene Expression

The function of CtBP1 and CtIP as corepressors of progesterone- or estrogen-regulated gene expression in the absence of LCoR was analyzed in MCF7 cells. The data raise the possibility that at least a portion of CtIP may be recruited to hormone-responsive promoters through its colocalization with LCoR in PRC1 complexes. Consistent with this hypothesis, we found that PRC1 marker BMI1 was also recruited to the pS2 promoter in the presence of estradiol (supplemental Fig. 1).

Deletion of the HTH Domain of LCoR Abolishes Corepressor Function—Function of the C-terminal HTH motif of LCoR in corepression of progesterone- and estrogen-regulated gene expression was analyzed by deletion of the domain (LCoRΔHTH; Fig. 5A) and comparison of the function of the resulting mutant to full-length LCoR. Western blotting and ChIP assays showed that the ΔHTH mutant was expressed at similar levels in T47D cells as full-length LCoR and that it was recruited to the IGFBP1 promoter in the presence of progesterone (Fig. 5, B and C). Remarkably, however, the LCoRΔHTH protein was essentially devoid of corepressor activity on either progesterone- or estrogen-responsive promoters in transient expression experiments (Fig. 5, D and E). In similar studies (not shown) corepressor activity of the LCoRΔHTH mutant was even more attenuated than that of LCoR derivative m1m2 lacking PXDLS motifs, identifying the HTH region as being critical to LCoR corepressor function.

Function of LCoR and CtBP1 as Attenuators of Progesterone-regulated Gene Expression—To further address the function of LCoR and CtBP1 in regulating hormone-dependent gene transcription, we knocked down expression of both proteins using siRNAs in T47D and MCF7 cells. Knockdown of cyclophilin B was used as the control for off-target effects of siRNA in these studies (Fig. 6, A–C). Loss of either LCoR or CtBP1 augmented progesterone-induced reporter gene expression in T47D cells (Fig. 6D) and estrogen-stimulated reporter expression in MCF7 cells (Fig. 6E), consistent with a role in corepression of hormone-inducible gene transcription.

We extended this analysis to the regulation in T47D cells of progesterone target genes encoding IGFBP1, mucin 1 (MUC1) and FK506-binding protein 5 (FKBP51) (4, 38, 39) (Fig. 7). Knockdown of LCoR markedly enhanced progesterone-stimulated expression of the IGFBP1 gene (Fig. 7A). Unexpectedly, ablation of CtBP1 expression alone or in combination with
LCoR Is a Corepressor of Progesterone-regulated Gene Expression

LCoR slightly attenuated basal and hormone-induced expression of the gene. In contrast, loss of LCoR or CtBP1 individually or in combination substantially augmented progesterone-stimulated expression of the mucin 1 (MUC1) and FK506-binding protein 5 (FKBP51) genes (Fig. 7, B and C). Taken together, the data in Figs. 6 and 7 provide evidence for roles of LCoR and CtBP1 as hormone-recruited attenuators of progesterone-regulated gene transcription. However, they also suggest that CtBP1 function may enhance the expression of some genes, similar to what was observed below in an analysis of estrogen-regulated gene expression.

Ablation of LCoR or CtBP1 Diminishes Expression of Estrogen Target Genes in a Gene-specific Manner—The effects of LCoR and/or CtBP1 knockdown on expression of a series of ERα target genes (40) were examined in MCF7 cells. Unlike the general stimulatory effect on progesterone-induced gene expression, knockdown of LCoR elicited distinct and gene-specific effects on estrogen target gene regulation. Notably, loss of CtBP1 but not LCoR attenuated estrogen-induced pS2 transcription (Fig. 8A). This effect of CtBP1 ablation is in agreement with previous studies showing that CtBP1 or CtBP2 knockdown attenuated estrogen-stimulated pS2 expression in MCF7 cells (41) and is consistent with the reduced expression of the IGFBP1 gene seen above in CtBP1-deficient cells. Although loss of CtBP1 had no effect on regulation of the GREB1 gene (the gene regulated by estrogen in breast cancer; Fig. 8B), its knockdown attenuated both basal and estrogen-regulated expression of the serum/glucocorticoids-regulated kinase 3 (SGK3) and cytochrome P450, family 26, subfamily b, polypeptide 1 (CYP26B1) genes (Fig. 8C and D). Knockdown of LCoR had no effect on basal or estrogen-induced expression of GREB1 gene (the gene regulated by estrogen in breast cancer; Fig. 8D), mimicking the

Downloaded from http://www.jbc.org/ by guest on November 18, 2013
Given the unexpected effects of knockdowns on estrogen-induced expression, we also analyzed the potential roles of LCoR and CtBP1 on genes whose transcription is repressed by...
LCoR Is a Corepressor of Progesterone-regulated Gene Expression

We have analyzed the roles of LCoR and its cofactor CtBP1 in coregulation of progesterone and estrogen receptor-regulated gene expression. Our previous findings showed that LCoR was a particularly efficacious inhibitor of progesterone-regulated reporter gene expression and that corepression was largely resistant to HDAC inhibitor trichostatin A (15). This study showed that CtBP1 did not repress hormone-regulated gene expression on its own in transient expression studies but did contribute to corepression by LCoR. GST pulldown experiments and communoprecipitations with a series of LCoR mutants suggested that the N-terminal region of LCoR containing the tandem PXDLS motifs was necessary and sufficient for interactions of CtBP1 with LCoR. Other experiments showed that deletion of the C-terminal HTH domain severely abrogated corepression, although this domain had no apparent role in CtBP1 recruitment. The HTH domain is distinct from the central region of LCoR identified in the accompanying manuscript (29) as being required for interaction with HDAC6. Together, these findings emphasize that LCoR is a multidomain protein that exerts its coregulator function through diverse mechanisms. siRNA-mediated knockdown of LCoR expression was enhanced on all three PR target genes tested in cells lacking LCoR.

The function of LCoR in corepression of PR-stimulated gene expression may be of considerable physiological importance. Progesterone signaling is essential for normal development and homeostasis of a number of physiological processes including female sexual behavior, ovulation, protection against seizures, maintenance of quiescence of the uterus during pregnancy, induction of germ cell maturation, and oocyte maturation (1, 5, 6). Progestins and anti-progestins are used clinically in contraception, hormone replacement therapy, induction of labor, treatment of endometriosis, and endometrial cancer (43). Moreover, expression of the PR along with ERα and human epidermal growth factor receptor 2 (HER2) are used as predictive markers for breast cancer therapy (44).

**DISCUSSION**

We further analyzed the molecular basis of these results by performing ChIP assays on the BMP7 promoter, where the ERα-binding region has been identified (42). Remarkably, we found that whereas LCoR was strongly recruited to the promoter after 30 min of estrogen treatment, CtBP1 partially dissociated from the promoter, indicating that the two factors function independently (Fig. 8G). ReChIP analysis found evidence for corecruitment of LCoR with ERα, but not CtBP1, on the BMP7 promoter after 30 min of estrogen treatment (not shown). These results are consistent with an effect of CtBP1 ablation on basal but not estrogen-regulated expression of the BMP7 gene.
There is considerable overlap in the expression patterns of LCoR and the PR. LCoR is widely expressed throughout development and in the adult (15). In tissue blots we observed the highest expression of LCoR in the placenta along with robust expression in several fetal tissues. Notably, the placenta is a site of progesterone biosynthesis, and in situ hybridization analysis...
LCOR Is a Corepressor of Progesterone-regulated Gene Expression

of near-term placenta (15) revealed that LCOR mRNAs were predominantly expressed in the syncytiotrophoblast layer of terminally differentiated cells, a site of PR expression and signaling (45, 46). The syncytiotrophoblast layer acts as a barrier between maternal circulation and the fetus, and its function is critical for controlling maternal signals that modulate fetal metabolism and development (47).

Compared with the substantial increases observed in progesterone-stimulated gene expression, the effects of LCOR or CtBP1 ablation on estrogen target genes were distinct and gene-specific. We observed no effect of LCOR knockdown on expression of three of four estrogen-stimulated genes studied. This may reflect a redundancy in corepressor function on the genes tested. For example, knockdown in MCF7 cells of NRP1, another corepressor recruited in the presence of hormone, had gene-specific effects on estrogen-dependent transactivation (48). LCOR ablation did augment both basal and hormone-stimulated expression of an estrogen-sensitive reporter gene, pointing to a potential role as an attenuator of ERα signaling. Remarkably, however, its knockdown blocked estrogen-stimulated expression of CYP26B1. This may reflect its function as a cofactor of CtBP1 on the CYP26B1 promoter, as ablation of CtBP1 also abolished estrogen-induced transcription. Notably, we found in the accompanying manuscript (29) that knockdown of HDAC6 had no effect on CYP26B1 regulation. A diminution in estrogen-stimulated gene expression in the absence of CtBP1 was also observed on two of three other estrogen-stimulated genes tested. Although these results were unexpected, they are consistent with other findings that knockdown of CtBP1 led to reduced expression of the ATP binding cassette, subfamily B (MDR/TAP) member 1 (MDR1) gene (49).

We also found that LCOR and CtBP1 are corecruited to the progesterone-stimulated IGFBP1 gene and the ERα target gene pS2. This behavior is in apparent contrast to the results of Stossi et al. (41), who found estrogen-induced dissociation of CtBP1 from the pS2 promoter. They found that CtBP1 controlled estrogen-mediated gene repression and that CtBP1 was recruited to repressed genes in the presence of estradiol. On the other hand, our observations are entirely consistent with other findings (50) in which peak recruitment of CtBP1 to the pS2 promoter was observed after 30 min of estradiol treatment.

CtBP1 and LCOR were extensively colocalized in the nucleus, including in pronounced foci. This pattern was similar to the colocalization of LCOR with CtIP and PRC1 marker BMI1 and strongly suggests that a substantial portion of CtBP1-bound LCOR is associated with PRC1 complexes. We found no evidence for direct interaction of LCOR with CtIP (Fig. 2) or BMI1 by GST pulldown assay. However, their indirect association is supported by coimmunoprecipitation of LCOR with both proteins and their hormone-dependent recruitment to estrogen or progesterone-regulated genes. Polycomb group proteins and complexes, including PRC1, form nuclear foci visualized as PcG (polycomb group proteins) bodies (51). Immunocytochemical studies have estimated these bodies to measure between 0.2 and 1.5 μm and to vary greatly in composition, suggesting that they represent foci with numerous independently functioning transcriptional regulatory complexes (25, 52).

More than 30 transcription factors have been shown to interact with CtBP1 (53). Recent studies characterized a CtBP-corepressor complex that contained a great number of proteins with opposing enzymatic activities. This included histone modifying enzymes (53) and other coregulatory proteins that can either activate or repress transcription depending on the context. Additionally, LCOR was identified as one of the components of a CtBP-corepressor complex purified from the nuclear extracts of HeLa cells (54). Taken together, these studies suggest that LCOR is associated with several distinct multisubunit transcriptional regulatory complexes; hence, implying the importance of LCOR in transcriptional control.

Acknowledgments—We are grateful to Jacynthe Laliberté for technical assistance with confocal microscopy. A special thank you goes to Dr. Myles Brown (Harvard Medical School) for providing the locations of EREs and ER-regulated enhancers.

REFERENCES