

# Tamoxifen reduces serum insulin-like growth factor I (IGF-I)

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## Summary

Antiestrogens are widely used in the management of hormonally responsive breast cancer in both adjuvant and palliative settings, and are currently being evaluated as chemopreventive agents. The classical mechanism of action of these drugs involves inhibition of estrogen-stimulated neoplastic cell proliferation by blockade of estrogen receptors present on breast cancer cells. This paper reviews recent clinical and laboratory data that suggest that the commonly used antiestrogen tamoxifen also acts to reduce serum IGF-I levels. Estrogens appear to play a permissive role in growth hormone (GH) release by the pituitary gland and GH is known to stimulate IGF-I expression by hepatocytes. It is therefore possible that blockade of estrogen receptors in the hypothalamic-pituitary axis by tamoxifen interferes with GH release, leading to reduced hepatic IGF-I expression. In view of results suggesting that IGF-I is a more potent mitogen than estradiol for breast cancer cells and data demonstrating a positive correlation between estrogen receptor level and IGF-I receptor level of breast cancer cells, the IGF-I lowering effect of tamoxifen may contribute to the cytostatic activity of the drug. The interrelationships between steroid hormone physiology and IGF-I physiology may have relevance to a variety of commonly used treatments for hormonally responsive cancers.

## Insulin-like growth factor I (IGF-I) responsiveness of breast cancer

A number of studies over the past decade provide support for the view that IGF-I [1] is an important mitogen for breast cancer, as well as certain other malignancies [2,3]. Studies of immortalized breast cancer cell lines in tissue culture demonstrate impressive stimulation of proliferation by IGF-I and inhibition of proliferation by blockade of type I IGF receptors [4-8]. In retrospect, the mitogenic responsiveness of breast cancer cells to

pharmacological concentrations of insulin demonstrated in older studies [9] is also consistent with an effect mediated by the type I IGF-I receptor. The remarkable inhibition of *in vivo* breast cancer proliferation resulting from blockade of IGF-I receptors [10] provides strong evidence to support the hypothesis that at least certain breast cancers are IGF-I responsive. Involvement of IGF-I in mammary cancer should come as no surprise given the role of this peptide in lactogenesis [11].

Studies of human breast cancer tissue have demonstrated specific IGF-I binding sites [12-17],

which have been localized by immunocytochemical studies to the neoplastic (as distinct from stromal) component of the tumour [18]. As there is a loose positive correlation between IGF-I receptor expression and estrogen receptor expression [13], it is likely that tumours that are identified by estrogen receptor assays as being hormonally responsive are influenced by both estrogens and by IGF-I. However, there also exists a class of estrogen receptor negative, IGF-I receptor positive human breast cancers.

### **Factors influencing tissue IGF-I bioactivity**

It is logical to propose that IGF-I-receptor positive breast cancer cells are influenced by tissue IGF-I bioactivity in their microenvironment rather than by serum IGF-I concentration. Quantitative measurements of IGF-I concentration or bioactivity are not available for human breast cancer tissue, but it is widely believed that local IGF-I bioactivity is influenced by factors which include both systemic and local production rates of both IGF-I and IGF-binding proteins. IGF-I binding proteins are a diverse group of circulating and tissue proteins that have high affinity binding sites for somatomedins, and modulate binding interactions between these growth factors and their receptors [1].

### *Control of IGF-I levels*

Factors controlling local production rates of IGF-I are incompletely described, but there is good evidence that the IGF-I gene is expressed by stromal cells of primary human breast cancers [19]. It is possible that stromal cells are capable of producing IGF-I not only in response to various physiological signals [20-24] (a stimulatory role for GH has not been excluded), but also in response to pathophysiological signals

originating from neighbouring tumour cells [19]. There is no strong evidence that autocrine production of IGF-I is a common event in human breast cancer.

More is known about the factors regulating serum IGF-I level, which is strongly influenced by hepatic production of IGF-I. Growth hormone is the chief regulator of IGF-I gene transcription in the liver [25,26], but permissive roles for thyroid and/or steroid hormones have not been excluded [27,28]. In view of pituitary influences on each of these hormones, hypothalamic homeostatic mechanisms ultimately play a key role in regulating serum IGF-I levels.

Neuroendocrine mechanisms may be involved in the negative correlation between age and serum IGF-I level, which has been noted in several studies [29]. Multiple pathways may be involved in the reduction of IGF-I levels associated with malnutrition [30]. While one may speculate that the negative relationship between serum IGF-I level and caloric intake is related in some way to the body of data associating Western diet with breast cancer incidence [31], such a straightforward relationship is probably simplistic, as the differences of intake associated with variability in breast cancer incidence are considerably less than the relatively dramatic reductions of intake that have been associated with reduced IGF-I levels in experimental systems [32].

In view of the key role played by growth hormone in regulating IGF-I gene expression, it is important to examine factors that determine pituitary growth hormone output. Secretion of growth hormone by pituitary somatotrophs is regulated by a complex neuroendocrine control system involving the stimulatory (growth hormone releasing factor) and inhibitory (somatostatin) hypothalamic releasing factors. These interact to produce a dynamic pulsatile pattern of growth hormone release [33]. The influences of steroid hormones on the secretion of GH are complex [34]. The possibility that these relationships might be involved in the action of various

commonly used endocrine therapies for neoplasia has received little attention until recently. There is evidence that estrogens play at least a permissive role in pituitary release of GH [26,35-40]. In addition, there is evidence that sex steroids influence the expression of somatostatin and/or growth hormone releasing factor in the hypothalamus [41-43]. Estrogen receptors that might mediate this process are indeed present in the hypothalamic-pituitary axis [44]. The relevance of these experimental studies to clinical medicine is underlined by studies demonstrating that administration of estrogens to individuals with hypogonadism who have low levels of estrogen and subnormal levels of GH and IGF-I can substantially correct abnormalities in the GH/IGF-I axis seen in this condition [45], a result consistent with the view that estrogens play a permissive role for GH secretion.

#### *Control of binding protein levels*

Our knowledge concerning the regulation of serum and tissue levels of the various IGF binding proteins remains incomplete, and is the subject of active investigation in many laboratories [1,46,47]. It is clear that somatomedin activity is influenced not only by the complex neuroendocrine systems that regulate GH secretion, but also by local concentrations of binding proteins. The existence of multiple levels of physiological control of somatomedin activity is in keeping with the view that this activity plays a crucial role in regulating cellular proliferation. Discussion of the proposed physiological roles and regulation of expression of the various IGF binding proteins is beyond the scope of this review, but it should be emphasized for those not working in the field that the binding proteins have affinity for IGFs in the same range as membrane receptors and are not 'non-specific' carrier proteins. Roles of estrogens and/or antiestrogens in regulating their expression have not been well

defined.

#### **Effect of tamoxifen on serum IGF-I levels — clinical studies**

In 1989, Colletti, Copeland, et al reported that mean serum IGF-I levels in a group of patients with metastatic breast cancer receiving tamoxifen was lower than that seen in a control group of breast cancer patients taking a variety of other antineoplastic treatments [48]. The patient population in this study was not uniform. Nevertheless, while the difference between groups was modest, it was statistically significant.

We took advantage of the availability of patients enrolled in a randomized, blinded, placebo-controlled clinical trial of tamoxifen in adjuvant therapy of breast cancer (NSABP B14 [49]) to independently test the hypothesis that antiestrogens might suppress the GH/IGF-I axis in humans by blockade of estrogen receptors in the hypothalamic-pituitary axis [50]. This study involved 69 patients, and yielded results similar to those reported by Colletti. Multivariate analysis confirmed that tamoxifen use was a significant predictor of serum IGF-I level, as was age. Although the reduction of mean IGF-I levels was modest (0.9 u/ml vs. 1.4 u/ml), the difference was statistically significant. Furthermore, the distribution of IGF-I levels was quite different between tamoxifen and placebo groups: in the former group 4.1% of individuals had IGF-I levels greater than 2 u/ml and 16.7% had levels less than 0.5 u/ml, as compared to 23.8% over 2 u/ml and 4.8% under 0.5 u/ml in the placebo-treated controls.

Recently, in collaboration with Craig Jordan and Andreas Fridel, we have extended this observation further by measuring serum IGF-I levels in a cohort of patients before and during tamoxifen therapy [51]. We observed a modest but statistically significant ( $p < .05$ ) decline in serum IGF-I level in individual patients following the

institution of tamoxifen therapy, a result consistent with the differences between groups taking placebo and tamoxifen in our previous study. In a small number of patients for whom multiple serum samples were available, we observed that the decline in IGF-I level associated with tamoxifen usage could persist for up to 5 years. Kiang et al [52] and Lien et al [53] have recently presented data which independently confirm the results of our randomized, placebo-controlled clinical study.

In all these studies, patient-to-patient heterogeneity with respect to the degree of suppression of IGF-I was noted. The lowest IGF-I levels were seen in post-menopausal women, who tended to have lower IGF-I levels to begin with, and to exhibit larger declines than younger women. Further clinical research now under way will attempt to determine whether there is any correlation between degree of decline of IGF-I and clinical response to treatment. While it is well known that older women tend to respond better to tamoxifen than younger women, the relationship between decline in IGF-I and response remains speculative at this time. Similarly, no data is available to test the hypothesis that relapse while on tamoxifen is related to escape from suppression of serum IGF-I level. Most work related to tamoxifen resistance has focused on events within neoplastic cells. While this line of research is promising, it does not exclude the possibility of 'host' factors contributing to tamoxifen resistance. There are a number of mechanisms by which such escape might be mediated, including increased GRH secretion and/or decreased somatostatin secretion. Further work is needed to evaluate this hypothesis.

### **Effect of tamoxifen on the GH/IGF-I axis — laboratory research**

Recent experimental work with primary cultures

of pituitary cells [54] provides support for the hypothesis that tamoxifen can directly inhibit pituitary growth hormone output. Similar experimental systems had previously been used to document the stimulatory and inhibitory influences of growth hormone releasing hormone (GRH) and somatostatin on somatotroph function [55]. We observed that clinically relevant concentrations of tamoxifen (1-10  $\mu$ M) significantly reduced the baseline production of GH, as assayed by radioimmunoassay of pituitary culture conditioned medium, and markedly attenuated the ability of the cultured cells to respond to GRH. The concentrations of tamoxifen employed did not affect the viability of the cultured pituitary cells. These results are consistent with the view that tamoxifen can inhibit GH synthesis and release at the level of the somatotroph. We speculate that this inhibition may involve pituitary estrogen receptors [54].

Chronically cannulated free-moving rats have been used for studies of growth hormone physiology, as *in vivo* GH secretion is pulsatile, and serum GH levels vary significantly from hour to hour. While this experimental system had previously been used to examine the effects of sex steroids on GH output [56], the effect of antagonists to sex steroids had not been studied. Tamoxifen, when administered in the manner originally described by Jordan as inhibiting DMBA-induced mammary cancer [57], had potent inhibitory effects on GH secretion, which persisted for at least 7 weeks following the administration of two doses of 5 mg subcutaneously [58]. This prolonged inhibition may well reflect the slow release and long half life of tamoxifen administered in this manner [59], rather than indicating a sustained inhibition following a transient exposure to the drug. Of considerable interest is the fact that the inhibitory effect of tamoxifen on the *in vivo* secretion of GH could be attenuated by an anti-somatostatin antiserum [58]. This implies that at least a part of the inhibitory effect of tamoxifen on GH output may be

mediated by increased release of endogenous somatostatin, a concept that is consistent with earlier reports showing that somatostatin gene expression in the rat hypothalamus can be regulated by sex steroids. Recent studies using tritiated estradiol have provided morphological evidence of estrogen receptors in both the arcuate-ventromedial and preoptic regions [60], the majority of which were localized to GRH secreting cells. Therefore, it is possible that over and above the direct effects of tamoxifen on pituitary cells implied by our *in vitro* studies, tamoxifen has a negative influence on GRH secretion and a positive influences on somatostatin secretion at the hypothalamic level. Furthermore, direct actions of tamoxifen on IGF-I gene expression by hepatocytes and/or other cell types have not been excluded.

## Discussion

### *Systemic effects of tamoxifen*

Although tamoxifen is a commonly used drug [61], our understanding of its mode or modes of action remains incomplete. Over the past decade, work from Lippman's group and others has provided important information concerning the sequelae of tamoxifen binding to estrogen receptors present on breast cancer cells. Examples of such events include stimulation of expression of the inhibitory factor TGF $\beta$  and inhibition of expression of the stimulatory factor TGF $\alpha$  [62,63]. Clinical and experimental research reviewed here provides evidence that tamoxifen has, in addition to its effects at the level of the breast cancer cell, an inhibitory effect on the GH/IGF-I axis. This property of tamoxifen is not, of course, the only systemic endocrine effect of the drug. There is evidence that tamoxifen suppresses estrogen-dependent prolactin secretion by the pituitary gland [64], and data to suggest that in premenopausal women, tamoxifen

can raise estrogen levels, presumably by blocking central estrogen receptors and interfering with feedback inhibition of gonadotrophin secretion [65].

While the characterization of a previously unrecognized effect of a drug as commonly used as tamoxifen is of interest, a key unanswered question concerns the relationship, if any, between suppression of the GH/IGF-I axis by tamoxifen and its antineoplastic properties. The degree of suppression of IGF-I levels associated with tamoxifen usage in humans is modest; it remains to be determined whether changes of this magnitude affect breast cancer proliferation. Co-administration of tamoxifen with compounds such as somatostatin analogues or GRH antagonists might be expected to suppress the GH/IGF-I axis more completely than tamoxifen alone [50,66,67]. A trial comparing the efficacy of tamoxifen with that of the combination of tamoxifen and a somatostatin analogue is underway at the Mayo Clinic, North Central Oncology Group, and McGill University.

This approach can be viewed as a step towards 'selective medical hypophysectomy'. By this, I refer to the pharmacological ablation of those pituitary functions which are non-essential to adult women, but which may nevertheless stimulate breast cancer proliferation. Candidate pituitary functions for inhibition include the GH/IGF-I axis, the LH/FSH/ovarian axis, and prolactin secretion, while ACTH and TSH are not targets. Recently, there has been interest in this concept that stems from the realization that growth hormone is an agonist for the prolactin receptor in humans [68]; thus, the older clinical trials [69] showing no effect of pharmacologically reducing prolactin secretion alone in human breast cancer were doomed to fail, even if lactogenic hormones do play a stimulatory role. This has led to at least one trial, currently under way, which is exploring the simultaneous pharmacological inhibition of prolactin and growth hormone secretion [70].

Research regarding the effect of tamoxifen on the GH/IGF-I axis now provides independent justification for further research in this area. Although there is significant species-to-species and individual-to-individual variability in central actions of tamoxifen, the drug clearly can, at least in certain experimental systems, reduce both prolactin [64] and growth hormone secretion [58], over and above its action at the estrogen receptors of neoplastic cells. Detailed quantification of these effects in humans, and information concerning their relevance to the antineoplastic actions of the drug, are unavailable. If the central actions of tamoxifen do contribute to the antineoplastic actions of the drug in humans, it is possible that enhancing these effects by combining tamoxifen with agents such as LHRH antagonists and/or somatostatin analogues will be of therapeutic value. It is also possible that variables related to pituitary function represent risk factors and/or prognostic factors for breast cancer.

Some older literature deserves re-examination in this context. Reports documenting responses of metastatic breast cancer to surgical hypophysectomy even in the setting of prior oophorectomy and adrenalectomy [71] are consistent with a role for the GH/IGF-I axis. Evidence consistent with the view that the stimulatory effect of estrogens on breast cancer cell proliferation involves the pituitary gland is found in classic studies showing that estrogens are incapable of stimulating *in vivo* proliferation of estrogen receptor positive DMBA-induced tumours in hypophysectomized rats [72]. While there is reason to interpret this result as evidence that estrogen-stimulated prolactin release by the pituitary is an important way in which estrogen stimulates neoplastic proliferation in the DMBA model [73], it should be recalled that DMBA-induced tumours are IGF-I-receptor positive [74], and this result is also consistent with a role for the GH/IGF-I axis in the *in vivo* proliferation of DMBA-induced tumours.

### *Multiple levels of interaction between estrogens and IGF-I*

There may be multiple levels at which estrogens (and antiestrogens) influence IGF-I physiology. In addition to the interactions emphasized in this review, there is laboratory evidence that expression of type I IGF-I receptors by ER positive breast cancer cells is up-regulated by exposure to estrogens [75], and down-regulated by antiestrogens [76]. Thus tamoxifen may not only reduce circulating IGF-I levels but also reduce responsiveness of the neoplastic cells to this mitogen, in addition to blocking any proliferation that is directly dependent on estrogens. This laboratory result is consistent with the observation that there is a positive correlation between estrogen receptor level and IGF-I receptor level in primary human breast cancer [13]. Those tumours with an ER negative, IGF-I-R positive phenotype may arise from abnormal regulation of IGF-I-R expression.

### *Role of circulating IGF-I in adults*

It is well known that patients who undergo surgical hypophysectomy for pituitary tumours require replacement of thyroid, adrenocortical, and often sex steroid hormones. In the adult, deficiency of GH and circulating IGF-I do not appear to be associated with major symptoms [77], although there is evidence that certain changes associated with aging may be related to the well described gradual decline of IGF-I levels with age [78]. In contrast, subnormal function of the GH-pituitary axis during growth and development is associated with well described pathology [79].

It would be fortuitous indeed if a subset of human malignancies exhibited a dependence on exogenous IGF-I, as novel endocrine therapies could exploit such a situation with minimal adverse effect on the patient. Perhaps tamoxifen as currently used acts in this way to a certain extent, not only in breast cancer, but also in certain desmoid [80] and pancreatic [81] tumours.

Neoplasms that are IGF-I-receptor negative and those that are IGF-I-receptor positive but express the IGF-I gene in an autocrine fashion would clearly not be candidates for this kind of treatment. A significant number of breast or other neoplasms [82], however, may respond to IGF-I from endocrine (circulating) or paracrine (e.g. stromal cell) sources. It is known that the former is growth hormone-dependent [83], and it will be important to extend previous work [20-24] to examine factors that regulate paracrine IGF-I expression, with particular attention to the role, if any, of growth hormone.

#### *Effects of tamoxifen on serum lipids*

A beneficial effect of tamoxifen on serum lipid levels has been described [84], and this effect may be of major clinical importance, particularly if ongoing trials of the drug lead to its widespread use as a preventative agent. The mechanism by which tamoxifen acts to alter lipid profiles remains incompletely characterized. As there is data to suggest that growth hormone raises serum triglyceride and cholesterol levels [85], it is possible that the growth hormone suppressive action of tamoxifen that we have reviewed here contributes to the lipid-lowering effects of the drug.

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#### **References**

1. Sara VR, Hall K: Insulin-like growth factors and their binding proteins. *Physiol Rev* 70:591-614, 1990

2. Macaulay VM: Insulin-like growth factors and cancer. *Br J Cancer* 65:311-320, 1992
3. Pollak MN, Polychronakos C, Richard M: IGF-I is a potent mitogen for human osteogenic sarcoma. *J Natl Cancer Inst* 82:301-305, 1990
4. Pollak MN, Polychronakos C, Yousefi S, et al: Characterization of insulin-like growth factor I (IGF-1) receptors of human breast cancer cells. *Biochem Biophys Res Commun* 149:276-281, 1988
5. Rohlik QT, Adams D, Kull FC Jr, et al: An antibody to the receptor for insulin-like growth factor I inhibits the growth of MCF-7 cells in tissue culture. *Biochem Biophys Res Commun* 149:276-281, 1987
6. Arteaga CL, Osborne CK: Growth inhibition of human breast cancer cells in vitro with an antibody against the type I somatomedin receptor. *Cancer Res* 49:6237-6241, 1989
7. Furlanetto R, DeCarlo J: Somatomedin-C receptors and growth effects in human breast cells maintained in long term tissue culture. *Cancer Res* 44:2122-2128, 1984
8. De Leon DD, Bakker B, Wilson DM, et al: Demonstration of insulin-like growth factor (IGF-1 and -II) receptors and binding protein in human breast cancer cell lines. *Biochem Biophys Res Commun* 152:398-405, 1988
9. Osborne CK, Bolan G, Monaco ME, Lippman ME: Hormone responsive human breast cancer in long term tissue culture: effect of insulin. *Proc Natl Acad Sci USA* 73:4536-4540, 1976
10. Arteaga CL, Kitten L, Coronado EB, et al: Blockade of the type I somatomedin receptor inhibits growth of human breast cancer cells in athymic mice. *J Clin Invest* 84:1418-1423, 1989
11. Duclos M, Houdebine LM, Djiane J: Comparison of insulin-like growth factor I and insulin effects on prolactin-induced lactogenesis in rabbit mammary gland in vitro. *Mol Cell Endocrinol* 65:129-134, 1989
12. Pollak MN, Perdue JF, Margolese RG, et al: Presence of somatomedin receptors on primary human breast and colon carcinomas. *Cancer Lett* 38:223-230, 1987
13. Pekonen F, Paranen S, Mäkinen T, et al: Receptors for epidermal growth factor and insulin-like growth factor I and their relation to steroid receptors in human breast cancer. *Cancer Res* 48:1343-1347, 1988
14. Peyrat JP, Bonneterre J, Beuscart R, et al: Insulin-like growth factor I receptors in human breast cancer and their relation to estradiol and progesterone receptors. *Cancer Res* 48:6429-6433, 1988
15. Berns EM, Klijn JG, van Staveren IL, Portengen H, Foekens JA: Sporadic amplification of the insulin-like growth factor I receptor gene in human breast tumors.

- Cancer Res 52:1036-1039, 1992
16. Foekens JA, Portengen H, van Putten WLJ, Trapman AMAC, Reubi JC, Alexieva-Figusch J, Klijn JGM: Prognostic value of receptors for insulin-like growth factor-1, somatostatin, and epidermal growth factor in human breast cancer. *Cancer Res* 49:7002-7009, 1989
  17. Foekens JA, van Putten WLJ, Portengen H, Rodenburg CJ, Reubi JC, Berns PMJJ, Henzen-Logmans S, van der Burg MEL, Alexieva-Figusch J, Klijn JGM: Prognostic value of pS2 protein and receptors for epidermal growth factor (EGF-R), insulin-like growth factor-1 (IGF-1-R), and somatostatin (SS-R) in patients with breast and ovarian cancer. *J Steroid Biochem Mol Biol* 37:815-821, 1990
  18. Pollak MN, Tremblay G: Immunocytochemical localization of insulin-like growth factor-I receptors in primary human breast cancers. *Breast Cancer Res Treat* 14:174, 1989
  19. Yee D, Paik S, Lebovic GS, Marcus RR, Favoni RE, Cullen KJ, Lippman ME, Rosen N: Analysis of insulin-like growth factor I gene expression in malignancy: Evidence for a paracrine role in human breast cancer. *Mol Endocrinol* 3:509-517, 1989
  20. D'Ercole AJ, Applewhite GT, Underwood LE: Evidence that somatomedin is synthesized by multiple tissues in the fetus. *Devel Biol* 75:315-328, 1980
  21. D'Ercole AJ, Stiles AD, Underwood LE: Tissue concentration of somatomedin C: Further evidence for multiple sites of synthesis and paracrine or autocrine mechanisms of action. *Proc Natl Acad Sci USA* 81:935-939, 1984
  22. Atkinson PR, Weidman ER, Bhaumick B, Bala RM: Release of somatomedin-like activity by cultured WI-38 human fibroblasts. *Endocrinology* 106:2006-2012, 1980
  23. Clemmons DR, Underwood LE, Van Wyk JJ: Hormonal control of immunoreactive somatomedin production by cultured fibroblasts. *J Clin Invest* 67:10-19, 1981
  24. Clemmons DR: Multiple hormones stimulate the production of somatomedin by cultured human fibroblasts. *J Clin Endocrinol Metab* 58:850-856, 1984
  25. Roberts CT, Brown AL, Graham DE, Seelig S, Berry S, Gabbay KH, Rechler MM: Growth hormone regulates the abundance of insulin-like growth factor I RNA in adult rat liver. *J Biol Chem* 261:10025-10028, 1986
  26. Mathews LS, Norstedt G, Palmiter RD: Regulation of insulin-like growth factor I gene expression by growth hormone. *Proc Natl Acad Sci USA* 83:9343-9347, 1986
  27. DiPippo VA, Powers CA: Estrogen induction of growth hormone in the thyroidectomized rat. *Endocrinology* 129:1696-1700, 1991
  28. Murphy LJ, Friesen HG: Differential effects of estrogen and growth hormone on uterine and hepatic insulin-like growth factor I gene expression in the ovariectomized hypophysectomized rat. *Endocrinology* 122:325-332, 1988
  29. Florini JR, Prinz PN, Vitiello MV, Hintz RL: Somatomedin-C levels in healthy young and old men: Relationship to peak and 24-hour integrated levels of growth hormone. *J Gerontol* 40:2-7, 1985
  30. Hintz R, Suskind R, Amatayakul K, Thanangkul O, Olson R: Plasma somatomedin and growth hormone values in children with protein-calorie malnutrition. *J Pediatr* 92:153-156, 1978
  31. Miller AB, Kelly A, Choi NW, Matthews V, Morgan RW, Munan L, Burch JD, Feather J, Howe GR, Jain M: A study of diet and breast cancer. *Am J Epidemiol* 107:499-509, 1978
  32. Straus DS, Takemoto CD: Effect of fasting on insulin-like growth factor-I (IGF-I) and growth hormone receptor mRNA levels and IGF-I gene transcription in rat liver. *Mol Endocrinol* 4:91-100, 1990
  33. Tannenbaum GS, Ling N: The interrelationship of growth hormone (GH)-releasing factor and somatostatin in generation of the ultradian rhythm of GH secretion. *Endocrinology* 115:1952-1957, 1984
  34. Jansson J-O, Eden S, Isaksson I: Sexual dimorphism in the control of growth hormone secretion in female rats. *Endocrinology* 121:1417-1423, 1987
  35. Simard J, Hubert J-F, Hosseinzadeh T, Labrie F: Stimulation of growth hormone release and synthesis by estrogens in rat anterior pituitary cells in culture. *Endocrinology* 119:2004-2011, 1986
  36. Wiedemann I, Schwartz E, Frantz AG: Acute and chronic estrogen effects upon serum somatomedin activity, growth hormone and prolactin in man. *J Clin Endocrinol Metab* 35:942-952, 1976
  37. Jin L, Song J, Lloyd RV: Estrogen stimulates both prolactin and growth hormone mRNA expression in the MtT/F4 transplantable pituitary tumor. *Proc Soc Exp Biol Med* 192:225-229, 1989
  38. Lloyd RV, Cano M, Landefeld TD: The effects of estrogens on tumor growth and on prolactin and growth hormone mRNA expression in rat pituitary tissues. *Am J Pathol* 133:397-406, 1988
  39. Shulman DI, Sweetland M, Duckett G, Root AW: Effect of estrogen on the growth hormone (GH) secretory response to GH-releasing factor in the castrate adult female rat *in vivo*. *Endocrinology* 120:1047-1051, 1987
  40. Ho KY, Evans WS, Blizzard RM, Veldhuis JD,

- Merriam GR, Samojlik E, Furlanetto R, Rogol AD, Kaiser DL, Thorner MO: Effects of sex and age on the 24-hour profile of growth hormone secretion in man: Importance of endogenous estradiol concentrations. *J Clin Endocrinol Metab* 64:51-58, 1987
41. Werner H, Koch Y, Baldino F Jr, Gozes I: Steroid regulation of somatostatin mRNA in the rat hypothalamus. *J Biol Chem* 263:7666-7671, 1988
42. Chowen-Breed JA, Steiner RA, Clifton DK: Sexual dimorphism and testosterone-dependent regulation of somatostatin gene expression in the periventricular nucleus of the rat brain. *Endocrinology* 125:357-362, 1989
43. Zeitler P, Argente J, Chowen-Breed JA, Clifton DK, Steiner RA: Growth hormone-releasing hormone messenger ribonucleic acid in the hypothalamus of the adult male rat is increased by testosterone. *Endocrinology* 127:1362-1368, 1990
44. McEwen BS, Davis PG, Parsons B, et al: The brain as a target for steroid hormone action. *Ann Rev Neurosci* 2:65-112, 1979
45. Cuttler L, Van Vliet G, Conte FA, et al: Somatomedin-C levels in children and adolescents with gonadal dysgenesis: Differences from age-matched normal females and effect of chronic estrogen replacement therapy. *J Clin Endocrinol Metab* 60:1087-1092, 1985
46. Minuto F, Barreca A, Del Monte P, Giordano G: Paracrine actions of IGF binding proteins. *Acta Endocrinologica (Copenh)* 124:63-69, 1991
47. Pollak M, Tetenes E, Maack C: Recombinant human insulin-like growth factor I binding protein 3 inhibits IGF-I-stimulated sarcoma cell proliferation [abs]. *Proc Am Assoc Cancer Res* 33:72, 1992
48. Colletti RB, Roberts JD, Devlin JT, et al: Effect of tamoxifen on plasma insulin-like growth factor I in patients with breast cancer. *Cancer Res* 49:1882-1884, 1989
49. Fisher B, Costantino J, Redmond C, et al: A randomized clinical trial evaluating tamoxifen in the treatment of patients with node-negative breast cancer who have estrogen-receptor-positive tumors. *N Engl J Med* 320:479-484, 1989
50. Pollak MN, Costantino J, Polychronakos C, Blauer S, Guyda H, Redmond C, Fisher B, Margolese R: Effect of tamoxifen on serum insulin-like growth factor I levels in stage I breast cancer patients. *J Natl Cancer Inst* 82:1693-1697, 1990
51. Friedl A, Jordan VC, Pollak M: Effect of tamoxifen on insulin-like growth factor I levels in breast cancer patients [abs]. *Proc Am Soc Clin Oncology* 11:52, 1992
52. Kiang DT, Kollander R, Kiang B, Kao PC: Role of plasma IGF-I in endocrine therapy for breast cancer [abs]. *Proc Am Soc Clin Oncology* 11:51, 1992
53. Lien EA, Johannessen DC, Aakvaag A, Lonning PE: Influence of tamoxifen, aminoglutethimide, and goserelin on human plasma IGF-I levels in breast cancer patients. *J Steroid Biochem Mol Biol* 41:541-543, 1992
54. Malaab SA, Pollak MN, Goodyer CG: Direct effects of tamoxifen on growth hormone secretion by pituitary cells *in vitro*. *Eur J Cancer*: in press, 1992
55. Blanchard MM, Goodyer CG, Charrier J, Barenton B: In vitro regulation of growth hormone (GH) release from ovine pituitary cells during fetal and neonatal development: Effects of GH-releasing factor, somatostatin, and insulin-like growth factor I. *Endocrinology* 122:2114-2120, 1988
56. Painsion J-C, Thorner MO, Krieg RJ, Tannenbaum GS: Short-term adult exposure to estradiol feminizes the male pattern of spontaneous and growth hormone-releasing factor-stimulated growth hormone secretion in the rat. *Endocrinology* 130:511-519, 1992
57. Jordan V: Effect of tamoxifen on initiation and growth of DMBA-induced rat mammary carcinomata. *Eur J Cancer* 12:419-424, 1976
58. Tannenbaum GS, Gurd W, Lapointe M, Pollak M: Tamoxifen attenuates pulsatile growth hormone secretion: Mediated in part by somatostatin. *Endocrinology* 130:3395-3402, 1992
59. Lien E, Solheim E, Ueland P: Distribution of tamoxifen and its metabolites in rat and human tissues during steady-state treatment. *Cancer Res* 51:4837-4844, 1991
60. Shirasu K, Stumpf WE, Sar M: Evidence for direct action of estradiol on growth hormone-releasing factor (GRF) in rat hypothalamus: Localization of [<sup>3</sup>H]estradiol in GRF neurons. *Endocrinology* 127:344-349, 1990
61. Lerner L, Jordan V: Development of antiestrogens and their use in breast cancer. *Cancer Res* 50:4177-4189, 1990
62. Knabbe C, Lippman M, Wakefield L, Flanders KC, Kasid A, Derynck R, Dixon RB: Evidence that transforming growth factor beta is a hormonally regulated negative growth factor in human breast cancer cells. *Cell* 48:417-428, 1987
63. Bates S, Davidson N, Valverius E, Freter CE, Dison RB, Tam JP, Kudlow JE, Lippman ME, Salomon DS: Expression of transforming growth factor alpha and its mRNA in human breast cancer: its regulation by estrogen and its possible functional significance. *Mol*

- Endocrinol 2:543-555, 1988
64. Jordan VC, Koerner S, Robison C: Inhibition of oestrogen-stimulated prolactin release by anti-oestrogens. *J Endocrinol* 65:151-152, 1975
  65. Groom GV, Griffiths K: Effect of the anti-oestrogen tamoxifen on plasma levels of luteinizing hormone, follicle-stimulating hormone, prolactin, oestradiol and progesterone in normal pre-menopausal women. *J Endocrinol* 70:421-428, 1976
  66. Pollak MN: Therapeutic implications of recent growth factor research. In Ragaz J, Ariel IM (eds) *High-Risk Breast Cancer*. Springer-Verlag, Berlin, 1991, pp 473-489
  67. Pollak MN, Polychronakos C, Guyda H: Somatostatin analogue SMS 201-995 reduces serum IGF-I levels in patients with neoplasms potentially dependent on IGF-I. *Anticancer Res* 9:889-891, 1989
  68. Fradkin JE, Eastman RC, Lesniak MA, Roth J: Specificity spillover at the hormone receptor — exploring its role in human disease. *N Engl J Med* 320:640-645, 1989
  69. Barrett A, Morgan L, Raggatt PR, Hobbs JR: Bromocriptine in the treatment of advanced breast cancer. *Clin Oncol* 2:373-377, 1976
  70. Pollak M, Poisson R, Major D, Gallant K: Octreotide with or without a prolactin suppressive agent for patients with breast cancer refractory to antiestrogens [abs]. *Proc Am Soc Clin Oncology* 11:87, 1992
  71. Ray BS, Pearson OH: Hypophysectomy in treatment of disseminated breast cancer. *Surg Clin North Am* 12:419-433, 1962
  72. Sterental A, Dominguez JM, Weissman C, Pearson OH: Pituitary role in the estrogen dependency of experimental mammary cancer. *Cancer Res* 23:481-485, 1962
  73. Arafah B, Manni A, Pearson O: Effect of hypophysectomy and hormone replacement on hormone receptor levels and the growth of 7,12-DMBA-induced mammary tumors in the rat. *Endocrinology* 107:1364-1369, 1980
  74. Ruggeri B, Klurfeld D, Kritchevsky D, Furlanetto R: Growth factor binding to 7,12-DMBA-induced mammary tumors from rats subject to caloric restriction. *Cancer Res* 49:4135-4141, 1989
  75. Stewart A, Johnson M, May F, Westley B: Role of insulin-like growth factors and the type I IGF receptor in the estrogen-stimulated proliferation of human breast cancer cells. *J Biol Chem* 265:21172-21178, 1990
  76. Freiss G, Rochefort H, Vignon F: Mechanisms of 4-hydroxytamoxifen anti-growth factor activity in breast cancer cells: Alteration of growth factor receptor binding sites and tyrosine kinase activity. *Biochem Biophys Res Commun* 173:919-926, 1990
  77. Anon: Growth hormone therapy in elderly people. *Lancet* 337:1131-1132, 1991
  78. Rudman D, Feller AG, Nagraj HS, Gergans GA, et al: Effects of human growth hormone in men over 60 years old. *N Engl J Med* 323:1-6, 1990
  79. Brock CGD: Growth hormone deficiency: Features assessment and therapy. In DeGroot LJ (ed) *Endocrinology*. Saunders, Philadelphia, 1989, pp 351-362
  80. Procter H, Singh L, Baum M, et al: Response of multicentric desmoid tumours to tamoxifen. *Br J Surg* 74:401, 1987
  81. Warshaw AL, Fernandez-del Castillo C: Medical progress: Pancreatic carcinoma. *N Engl J Med* 326:455-465, 1992
  82. Pollak MN, Sen A, Richard M, Tetenes E, Bell R: Inhibition of metastatic behavior of murine osteosarcoma by hypophysectomy. *J Natl Cancer Inst* 84: in press, 1992
  83. Schwander JC, Hauri C, Zapf J, Froesck ER: Synthesis and secretion of insulin-like growth factor and its binding protein by the perfused rat liver: Dependence on growth hormone status. *Endocrinology* 113:297-305, 1983
  84. Love R, Newcomb P, Wiebe D, Surawicz T, Jordan VC, Carbone PP, Demets DL: Effects of tamoxifen therapy on lipid and lipoprotein levels in postmenopausal patients with node-negative breast cancer. *J Natl Cancer Inst* 82:1327-1332, 1990
  85. Moller A, Rasmussen LM, Thuesen L, Christiansen JS: Impact of human growth hormone on plasma lipoprotein concentrations. *Horm Metabol Res* 21:207-209, 1989