

**LRRK2-KO/AIW002-02**

Control line  Disease line  
Gene edited? Yes

**General information**

Cell line name	LRRK2-KO/AIW002-02 No. 2-2-2
Biosample ID	2889
Lines from same donor	

**Donor information**

Sex	MALE
Age	37 YEARS
Race	CAUCASIAN

**Culture conditions**

Coating/medium	MATRIGEL/mTeSR1
Passage method	Gentle cell dissociation reagent

**Derivation**

Primary cell line	PBMC
Reprogramming method	RETROVIRUS
Reprogramming factors	Bcl-XI <input type="checkbox"/> Myc <input checked="" type="checkbox"/> Nanog <input type="checkbox"/> SOX2 <input checked="" type="checkbox"/> LIN28 <input type="checkbox"/> KLF4 <input checked="" type="checkbox"/> OCT3/4 <input checked="" type="checkbox"/>

**Disease status**

Disease	Parkinson Disease
Affected gene	LRRK2
Disease mutation/family history	LRRK2-KO

**Genetic modification**

Modification	CRISPR-Cas9 Knock out
Gene	LRRK2
Gene ID	ENSG00000188906
Chromosome location	Chromosome 12: 40,196,744-40,369,285
gRNA1	gRNA1: CAACAGCTGGAGATGGCGGT,
gRNA2	gRNA2: GTCCTAGAGTAAGGCACATT.
Delivery method	Lipofectamine TM stem reagent
Subclone IDs	No.2-2-2, No.2-5-14

**Description**

For knocking out the expression of LRRK2 in AIW002-02, guide RNAs (gRNAs) were designed using the “optimized CRISPR design” tool ([www.crisp.mit.edu](http://www.crisp.mit.edu)). The locations of gRNA1 and gRNA2 were around exon 1 with ATG start codon. Oligonucleotides with Bsb1 cleavage overhang were ordered from Life Technologies, annealed and cloned into Cas9/puromycin expressing vector (PX459 from Addgene #48139). The following gRNA sequences were used to create the KO line. gRNA1: CAACAGCTGGAGATGGCGGT, gRNA2: GTCCTAGAGTAAGGCACATT.

Genotyping: genomic DNA was extracted with QuickExtract (Lucigen) and PCR was performed using Q5® High-Fidelity DNA Polymerase according to the manufacturer’s protocol (Forward primer: TTCCTCATAAACAGGCGGGC . Reverse primer:

AAGAGTCCGGGTTTGAGCAG). Wild type PCR product: 427bp. Knock out PCR product: 320bp.

LRRK2 KO/AIW002-02 No. 2-2-2

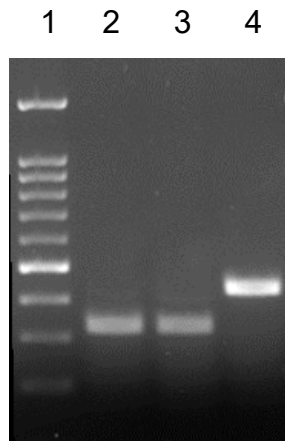
One chromosome, deletion 106 bases result in reading frame shift

```
GCCCCCGGGGAGCTGTGGCCGGCGCCCTGCCGGTTCCTGAGCA
GCGGACGTTTCATGCTGGGAGGGCGGCGGGTGGAAAGCAGGTGCCAC
CATGGCATCCTGGAGGATCTGCTGGTGTTCACGTACTCCGAGCGCGG
TAATCACTTGAAAATAAACTGTGCTTTTATTTTT
```

The other chromosome, deletion 107 bases result in reading frame shift.

```
GCCCCTGCCGGTTCCTGAGCAGCGGACGTTTCATGCTGGGAGGGCG
GCGGGTGGAAAGCAGGTGCCACCATGGCTCCTGGAGGATCTGCTGGT
GTTACGTACTCCGAGCGCGGTAATCACTTGAAAATAAACTGTGCTTTT
ATTTTTG
```

1A



1. 100bp DNA ladder
2. LRRK2-KO/AIW002-02 No. 2-2-2
3. LRRK2-KO/AIW002-02 No. 2-5-14
4. AIW002-02

1B

```
ACCATGGCTAGTGGCAGCTGTCAAGGGGTCGAAAGAGGACGAGGAAACTCTGAAGAAGTTGATAGTCAGGCTGAACAATGTCCAGGAAAGGAAAACAGATAGAAAACGCTGGTCCAAATCCTGGAGGATCT
ACCATGGC-----ATCCTGGAGGATCT
ACCATGGCT-----CCTGGAGGATCT
```

**Figure 1. Sequence verification of LRRK2 knock-out in AIW002-02.**

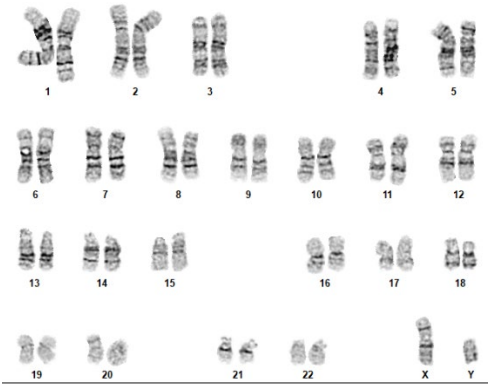
A. PCR analysis of LRRK2-KO/AIW002-02: 320bp. AIW002-2: 427bp

B. Sequences of LRRK2-KO/AIW002-02 is aligned with AIW002-02 sequence. Top: AIW002-02 genomic DNA sequence. Bottom: two chromosome of LRRK2-KO/AIW002-02 No.2-2-2.

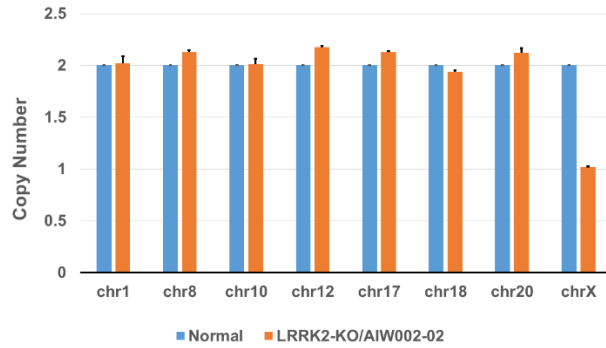
### Characterization:

**Genotyping analysis:**

Passage number	P12+P6 No.2-2-2
Karyotyping	Normal 46,XY
qPCR (Genetic Analysis kit)	Normal



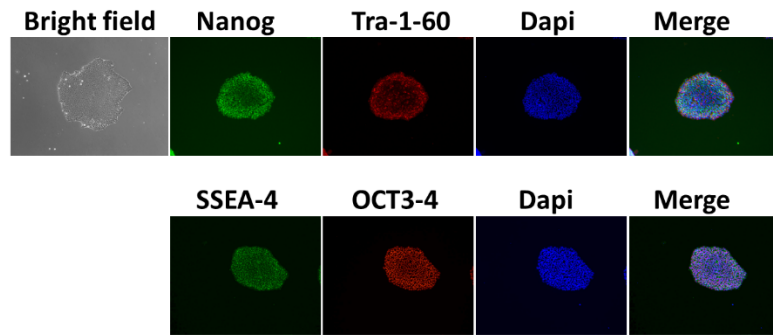
**Figure 2.**G-band assay show normal karyotype of LRRK2-KO/AIW002-02 No. 2-2-2, 46, XY



**Figure 3.** Genetic stability assay show normal chromosome of LRRK2-KO/AIW002-02 No. 2-2-2

**Pluripotency analysis:**

Marker	Expressed?
Nanog	Yes
Tra-1-60	Yes
SSEA-4	Yes
OCT3/4	Yes



**Figure 4.** Bright field image and immunostaining of pluripotency markers on LRRK2-KO/AIW002-02 No.2-2-2.

**Microbiology/virus screening:**

Mycoplasma test	Negative
Hepatitis B	Negative
Hepatitis C	Negative
HIV1 or 2	Negative